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Product Information

MISSION® shRNA Plasmid DNA

Catalog Number **SHCLND** Storage Temperature –20 °C

TECHNICAL BULLETIN

Product Description

Small interfering RNAs (siRNAs) processed from short hairpin RNAs (shRNAs) are a powerful way to mediate gene specific RNA interference (RNAi) in mammalian cells. The MISSION product line is a viral-vector-based RNAi library against annotated mouse and human genes. shRNAs that are processed into siRNAs intracellularly are expressed from amphotropic lentivirus particles, allowing screening in a wide range of mammalian cell lines. In these cell lines, MISSION shRNA clones permit rapid, cost-efficient loss-of-function and genetic interaction screens.

The TRC1 and TRC1.5 libraries consist of sequence-verified shRNA lentiviral plasmid vectors for mouse and human genes cloned into the pLKO.1-puro vector (see Figure 1). The TRC2 library consists of sequence-verified shRNAs for mouse and human genes in the TRC2-pLKO-puro vector (see Figure 2). The TRC2 vector has a single additional element in comparison to the TRC1 vector. This is the WPRE,¹ or the Woodchuck Hepatitis Post-Transcriptional Regulatory Element. WPRE allows for enhanced expression of transgenes delivered by lentiviral vectors.²

A number of individual shRNAs designed using a proprietary algorithm are available for each gene. We recommend purchasing multiple individual constructs (the recommended number is listed on each clone ordering page) targeting different regions of the gene sequence.

Target cell lines may be transfected with the purified plasmid for transient or stable gene silencing (puromycin selection). In addition, self-inactivating replication incompetent viral particles can be produced in packaging cells (HEK293T) by co-transfection with compatible packaging plasmids.⁴⁻⁵ Unlike murine-based MMLV or MSCV retroviral systems, lentiviral-based particles permit efficient infection and

integration of the specific shRNA construct into differentiated and non-dividing cells, such as neurons and dendritic cells, overcoming low transfection and integration difficulties when using these cell lines.

Each MISSION shRNA clone is constructed within the lentivirus plasmid vector pLKO.1-puro³ or TRC2-pLKO-puro. Both vectors contain the ampicillin and puromycin antibiotic resistance genes for selection of inserts in bacterial or mammalian cells respectively.

A range of knockdown efficiencies can be expected when using multiple clones. This allows one to examine the effect of loss of gene function over a large series of gene knockdown efficiencies. Each shRNA construct has been cloned and sequence verified to ensure a match to the target gene.

Components/Reagents

The individual constructs are provided as frozen stocks containing 1 μ g of plasmid DNA in Tris-EDTA (TE) buffer at a concentration of 20 ng/ μ L.

Orders of 25 or fewer clones are provided in individual vials. Each vial contains a unique one dimensional barcode label that can be read using a corresponding reader. A printed value corresponding to The RNAi Consortium (TRC) clone number is also provided on each tube. Orders of >25 clones are provided in a 96-well plate with a one dimensional barcode label on the plate. 96-well plates are provided with a CD containing plate map positions.

The hairpin sequence, other unique clone information and additional gene related products including antibodies and small molecules can be found through our comprehensive search tool, Your Favorite Gene www.sigma-aldrich.com/yfg using RefSeq accession numbers, e.g., NM_027088, unique clone identification numbers, e.g., NM_027088.1-989s1c1, or TRC numbers, e.g., TRCN0000030720.

Precautions and Disclaimer

These products are for R&D use only, not for drug, household, or other uses. Please consult the Material Safety Data Sheet for information regarding hazards and safe handling practices.

Figure 1. TRC1 and TRC1.5 Lentiviral Plasmid Vector pLKO.1-puro Features

vector percon-puro reatures						
Name	Description					
U6	U6 Promoter					
cppt	Central polypurine tract					
hPGK	Human phosphoglycerate kinase					
	eukaryotic promoter					
puroR	Puromycin resistance gene for					
	mammalian selection					
SIN/3'	3' self inactivating long terminal					
LTR	repeat					
f1 ori	f1 origin of replication					
ampR	Ampicillin resistance gene for					
	bacterial selection					
pUC ori	pUC origin of replication					
5' LTR	5' long terminal repeat					
Psi	RNA packaging signal					
RRE	Rev response element					

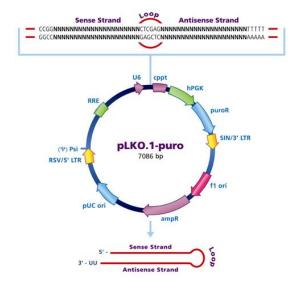
Figure 2. TRC2 Lentiviral Plasmid Vector TRC2pLKO-puro Features

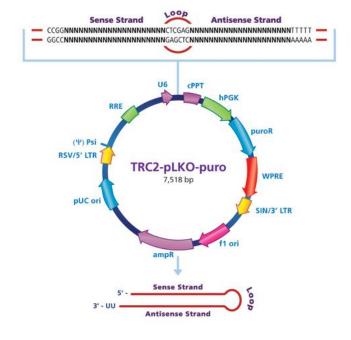
perto-puro i catures						
Name	Description					
U6	U6 Promoter					
cppt	Central polypurine tract					
hPGK	Human phosphoglycerate kinase eukaryotic promoter					
puroR	Puromycin resistance gene for mammalian selection					
WPRE	Woodchuck Hepatitis Post- Transcriptional Regulatory Element					
SIN/3'	3' self inactivating long terminal					
LTR	repeat					
f1 ori	f1 origin of replication					
ampR	Ampicillin resistance gene for bacterial selection					
pUC ori	pUC origin of replication					
5' LTR	5' long terminal repeat					
Psi	RNA packaging signal					
RRE	Rev response element					

Storage/Stability

All components are stable for at least one year after receipt when stored at $-20~^{\circ}C$.

Gently spin down material in tubes before opening.





Control Selection Table

Sigma's recommended controls for any shRNA experiment are closely aligned with the controls suggested in the *Nature Cell Biology* editorial. Additional controls are available. Please visit: http://www.sigmaaldrich.com/life-science/functional-genomics-and-rnai/shrna/trc-shrna-products/shrna-controls.html

Recommended Control	Objective				
Negative Control: Untreated Cells	Untreated cells will provide a reference point for comparing all other samples.				
Negative Control: Transfection with empty vector, containing no shRNA insert	MISSION pLKO.1-puro Control Vector, Catalog Number SHC001 The empty vector, pLKO.1-puro, is a useful negative control that will not activate the RNAi pathway because it does not contain an shRNA insert. It will allow for observation of cellular effects of the transfection process and the delivery of the lentiviral vector. Cells transfected with the empty vector provide a useful reference point for comparing specific knockdown.				
Negative Control: Transfection with non- targeting shRNA	MISSION Non-Target shRNA Control Vector, Catalog Number SHC002 This non-targeting shRNA vector is a useful negative control that will activate RISC and the RNAi pathway, but does not target any human or mouse genes. The short-hairpin sequence contains 5 base pair mismatches to any known human or mouse gene. This allows for examination of the effects of shRNA transfection on gene expression. Cells transfected with the non-target shRNA vector will also provide a useful reference for interpretation of knockdown.				
Positive Control: Transfection with positive reporter vector	MISSION TurboGFP™ Control Vector, Catalog Number SHC003 This vector is a useful positive control for measuring transfection efficiency and optimizing shRNA delivery. The TurboGFP Control vector consists of the lentiviral backbone vector, pLKO.1-puro, containing a gene encoding TurboGFP, driven by the CMV promoter. Transfection of this vector provides fast visual confirmation of successful transfection and delivery.				
Positive Control: Transfection with shRNA targeting reporter vector	MISSION TurboGFP shRNA Control Vector, Catalog Number SHC004 The TurboGFP shRNA vector consists of the pLKO.1–puro vector, containing shRNA that targets TurboGFP, and can be used as a positive control to quickly visualize knockdown. This TurboGFP shRNA Control Vector has been experimentally shown to reduce GFP expression by 99.6% in HEK 293T cells after 24 hours. Because this vector targets TurboGFP, and it does not target any human or mouse genes, it can also be used as a negative non-target control in shRNA experiments.				

Cell Type Table

The cell types listed below have been successfully infected by pLKO.1-puro based shRNA constructs. Optimal conditions will need to be determined for your experimental needs. For the most updated cell line list, and some guidelines for conditions, please visit:

http://www.sigmaaldrich.com/life-science/functional-genomics-and-rnai/shrna/learning-center/getting-started.html

Cell lines, human	Cell Type	Cell lines, human	Cell Type	Primary cells human	Cell Type
HEK293	embryonic kidney cells	A431	epidermal carcinoma	dendritic	immature dendritic
HeLa	cervical adenocarcinoma	THP1	monocytic	T-cells	lymphocytes
A549	lung adenocarcinoma	RAW264.7	macrophage	epithelial	prostate
H1299	lung carcinoma	SH-SY5Y	brain neuroblastoma	fibroblasts	primary mammary
HT29-D4	colon carcinoma	HCN-1A	brain cortical neuron	Primary cells, other species	Cell Type
HepG2	hepatocellular carcinoma	SupT1	T-cells	ECS	mouse embryonic stem cells
HCT116	colon carcinoma	BJ-TERT	diploid fibroblasts	fibroblasts	mouse embryonic fibroblasts
MCF7	breast carcinoma	Cell lines, mouse	Cell Type	MC3T3-E1	mouse bone marrow derived
MCF10A	breast carcinoma	NIH3T3	fibroblast	molar mesenchymal	mouse embryonic mesenchymal
Panc-1	pancreatic epithelioid carcinoma	Primary cells, human	Cell Type	cardiomyocytes	rat neonatal cardiomyocytes
PC3	prostate carcinoma	astrocytes	normal		
DU145	prostate carcinoma	C3H10T1/2	mesenchymal		

References

- Donello, J.E., et al., Woodchuck hepatitis virus contains a tripartite posttranscriptional regulatory element. J Virol., 72, 5085-5092 (1998).
- 2. Zufferey, R., et al., Woodchuck hepatitis virus posttranscriptional regulatory element enhances expression of transgenes delivered by retroviral vectors. *J Virol.*, 73, 2886-2892 (1999).
- 3. Stewart, S.A., *et al.*, Lentivirus-delivered stable gene silencing by RNAi in primary cells. *RNA*, 9, 493-501 (2003).
- 4. Zufferey, R., et al., Multiply attenuated lentiviral vector achieves efficient gene delivery in vivo. Nat. Biotechnol. 15, 871-885 (1997).
- 5. Zufferey, R., *et al.*, Self-inactivating lentivirus vector for safe and efficient *in vivo* gene delivery. *J Virol.*, 72, 9873-9880 (1998).
- 6. Whither RNAi? *Nature Cell Biology*, 5, 489-490 (2003).

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U.S. Patent No. 6,136,597, U.S. Patent No. 6,284,469, U.S. Patent No. 6,312,912, U.S. Patent No. 6,287,814.

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Fax: (858) 546-8093

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