

User Guide

MILLIPLEX® TGFβ1 Single Plex Magnetic Bead Kit

96-Well Plate Assay

TGFBMAG-64K-01
TGFBMAG-64K-01BK

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Introduction

The transforming growth factor beta (TGF β) system, a superfamily of cytokines as well as signaling pathways, is highly conserved throughout the animal kingdom. TGF β functions in angiogenesis, wound healing and embryonic development, and plays a critical role in immunity, heart disease, and cancer. In its normal state TGF β is one of the few classes of proteins able to inhibit cell growth by halting mitosis at the G1 state, inducing cell differentiation or apoptosis. However, during oncogenesis mutations in the TGF β signaling pathway result in tumor cell resistance to the effects of normally functioning TGF β , causing proliferation without regulation. Initial research suggests that VE-cadherin may enhance the mutated TGF β signaling pathway, while other research indicates that DNA methylation plays a role in pathway mutation.

The secreted TGF β cytokine exists in three isoforms: TGF β 1, TGF β 2 and TGF β 3. Secreted by most immune cells, TGF β 1 plays a critical role in controlling the immune system, acting on cells differently depending on cell type as well as stage of differentiation. TGF β 2, also known as glioblastoma-derived T-cell suppressor factor (G-TSF), plays a role in embryonic development and has the ability to suppress the effects of interleukin-dependent T-cell tumors. TGF β 3 regulates cellular adhesion molecules and extracellular matrix formation, as well as lung and palate development. TGF β 3 deficiency during mammalian development results in the cleft palate deformity. In addition, TGF β 3 controls wound healing by regulating epidermal and dermal cell movement in injured skin.

The MILLIPLEX[®] portfolio offers the broadest selection of analytes across a wide range of disease states and species. Once the analytes of interest have been identified, you can rely on the quality that we build into each kit to produce results you can trust. In addition to the assay characteristics listed in the protocol, other performance criteria evaluated during the verification process include: cross-reactivity, dilution linearity, kit stability, and sample behavior (for example, detectability and stability).

Each MILLIPLEX[®] panel and kit includes:

- Quality controls (QCs) provided to qualify assay performance
- Comparison of standard (calibrator) and QC lots to a reference lot to ensure lot-to-lot consistency
- Optimized serum matrix to mimic native analyte environment
- Detection antibody cocktails designed to yield consistent analyte profiles within panel

In addition, each panel and kit meets stringent manufacturing criteria to ensure batch-to-batch reproducibility. The MILLIPLEX[®] TGF β 1 Single Plex Magnetic Bead Kit thus enables you to focus on the therapeutic potential of TGF β protein signaling pathways. Coupled with the Luminex[®] xMAP[®] platform in a magnetic bead format, you receive the advantage of ideal speed and sensitivity, allowing quantitative multiplex detection of dozens of analytes simultaneously, which can dramatically improve productivity.

The MILLIPLEX® TGFβ1 Single Plex Magnetic Bead Kit is part of the most versatile system available for immunology research. From our single to multiplex biomarker solutions, we partner with you to design, develop, analytically verify and build the most comprehensive library available for protein detection and quantitation. MILLIPLEX® products offer you a convenient “all-in-one” box format that gives you the assurance that you will have all the necessary reagents you need to run your assay.

The MILLIPLEX® TGFβ1 Single Plex Magnetic Bead Kit is a single plex kit to be used for the quantification of TGFβ in serum, plasma, tissue/cell lysate and culture supernatant samples.

**For research use only. Not for use in diagnostic procedures.
Please read entire protocol before use.
It is important to use same assay incubation conditions throughout your study.**

Principle

MILLIPLEX® products are based on the Luminex® xMAP® technology — one of the fastest growing and most respected multiplex technologies offering applications throughout the life-sciences and capable of performing a variety of bioassays including immunoassays on the surface of fluorescent-coded magnetic bead (MagPlex®-C and non-magnetic bead (MicroPlex®) microspheres

- Luminex® products use proprietary techniques to internally color-code microspheres with multiple fluorescent dyes. Through precise concentrations of these dyes, distinctly colored bead sets of 500-5.6 μm non-magnetic or 80-6.45 μm magnetic polystyrene microspheres can be created, each of which is coated with a specific capture antibody.
- After an analyte from a test sample is captured by the bead, a biotinylated detection antibody is introduced.
- The reaction mixture is then incubated with Streptavidin-PE conjugate, the reporter molecule, to complete the reaction on the surface of each microsphere.
- The following Luminex® instruments can be used to acquire and analyze data using two detection methods:
 - The Luminex® analyzers, Luminex® 200™, FLEXMAP 3D®, and xMAP® INTELLIFLEX, are flow cytometry-based instruments that integrate key xMAP® detection components, such as lasers, optics, advanced fluidics and high-speed digital signal processors.
 - The Luminex® analyzer (MAGPIX®), a CCD-based instrument that integrates key xMAP® capture and detection components with the speed and efficiency of magnetic beads.

-
- Each individual microsphere is identified and the result of its bioassay is quantified based on fluorescent reporter signals. We combine the streamlined data acquisition power of Luminex® xPONENT® acquisition software with sophisticated analysis capabilities of the new MILLIPLEX® Analyst 5.1, integrating data acquisition and analysis seamlessly with all Luminex® instruments.
 - xMAP® INTELLIFLEX runs on INTELLIFLEX software for instrument control, run setup and generating high quality data with flexible output options. Data can be exported in xPONENT® style CSV files for compatibility with many existing analytical applications, or in the new, customizable INTELLIFLEX file format. The INTELLIFLEX file format is intended for flexibility and simplicity, allowing the user to freely select which data points to include and to reduce the time to analysis.

The capability of adding multiple conjugated beads to each sample results in the ability to obtain multiple results from each sample. Open-architecture xMAP® technology enables multiplexing of many types of bioassays reducing time, labor and costs over traditional methods.

Storage Conditions Upon Receipt

- Recommended storage for kit components is 2–8 °C.
- For long-term storage, freeze reconstituted standards and controls at ≤ -20 °C. Avoid multiple (> 2) freeze/thaw cycles.
- **DO NOT FREEZE** Antibody-Immobilized Beads, Detection Antibody, and Streptavidin-Phycoerythrin.

Reagents Supplied

Store all reagents at 2–8 °C.

| Reagents Supplied | Volume | Quantity | Cat. No. |
|--|-------------|-----------|-----------|
| TGFβ1 Standard | Lyophilized | 1 vial | LTGF-8064 |
| TGFβ1 Quality Controls 1 and 2 | Lyophilized | 2 vials | LTGF-6064 |
| TGFβ1 Detection Antibody | 3.2 mL | 1 bottle | LTGF-1064 |
| Streptavidin-Phycoerythrin | 3.2 mL | 1 bottle | L-SAPE9 |
| Serum Matrix | Lyophilized | 1 vial | LMC-SD |
| Sample Diluent | 5.0 mL | 1 bottle | LTGF-SD |
| 1.0 N Hydrochloric Acid (HCl) | 1.0 mL | 1 bottle | L-HCL |
| 1.0 N Sodium Hydroxide (NaOH) | 1.0 mL | 1 bottle | L-NAOH |
| Assay Buffer | 30 mL | 2 bottles | L-AB1 |
| 10X Wash Buffer | 60 mL | 1 bottle | L-WB |
| Set of one 96-Well Microtiter Plate with 2 Sealers | - | 1 set | - |

TGFβ1 Single Plex Antibody-Immobilized Magnetic Beads

| Bead/Analyte Name | Luminex® Magnetic Bead Region | 1X concentration, 3.5 mL | |
|-------------------|-------------------------------|--------------------------|-----------|
| | | Available | Cat. No. |
| Anti-TGFβ1 Bead | 18 | ✓ | TGFB1-MAG |

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Materials Required (Not included)

Reagents

MAGPIX® Drive Fluid PLUS (Cat. No. 40-50030), xMAP® Sheath Fluid PLUS (Cat. No. 40-50021), or xMAP® Sheath Concentrate PLUS (Cat. No. 40-50023)

Instrumentation/Materials






- Adjustable pipettes with tips capable of delivering 25 µL to 1000 µL
- Multichannel pipettes capable of delivering 5 µL to 50 µL, or 25 µL to 200 µL
- Reagent reservoirs
- Polypropylene microfuge tubes
- Rubber bands
- Aluminum foil
- Absorbent pads
- Laboratory vortex mixer
- Sonicator (Branson Ultrasonic Cleaner Model No. B200 or equivalent)
- Titer plate shaker (VWR® Microplate Shaker Cat. No. 12620-926 or equivalent)
- Luminex® 200™, HTS, FLEXMAP 3D®, MAGPIX® instrument with xPONENT® software, or xMAP® INTELLIFLEX instrument with INTELLIFLEX software by Luminex® Corporation
- Automatic Plate washer for magnetic beads (BioTek® 405 LS and 405 TS, Cat. No. 40-094, 40-095, 40-096, 40-097 or equivalent) or Handheld Magnetic Separation Block (Cat. No. 40-285 or equivalent).

Note: If a plate washer or handheld magnetic separation block for magnetic beads is not available, one can use a microtiter filter plate (Cat. No. MX-PLATE) to run the assay using a vacuum filtration unit (Vacuum Manifold, Cat. No. MSVMHTS00 or equivalent with Vacuum Pump, Cat. No. WP6111560 or equivalent).

Safety Precautions

- All blood components and biological materials should be handled as potentially hazardous. Follow universal precautions as established by the Centers for Disease Control and Prevention and by the Occupational Safety and Health Administration when handling and disposing of infectious agents.
- Sodium azide has been added to some reagents as a preservative. Although the concentrations are low, Sodium azide may react with lead and copper plumbing to form highly explosive metal azides. Dispose of unused contents and waste in accordance with international, federal, state, and local regulations.

Symbol Definitions

| Ingredient | Cat No. | Label | |
|-------------------------------|---------|---|---|
| 1.0 N NaOH | L-NAOH |  | Danger. May be corrosive to metals. Causes severe skin burns and eye damage. Wear protective gloves/protective clothing/eye protection/face protection. IF SWALLOWED: Rinse mouth. Do NOT induce vomiting. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. IF exposed or concerned: immediately call a POISON CENTER or doctor/physician. |
| Serum Matrix | LMC-SD | No Symbol Required | Harmful to aquatic life with long lasting effects. Avoid release to the environment. |
| Streptavidin-Phycoerythrin | L-SAPE9 |  | Warning. Causes serious eye irritation. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. |
| Wash Buffer | L-WB |  | Warning. May cause an allergic skin reaction. Wear protective gloves. IF ON SKIN: Wash with plenty of soap and water. |
| Assay Buffer | L-AB1 |  | Warning. Causes serious eye irritation. IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. |
| 1.0 N Hydrochloric Acid (HCl) | L-HCL |  | Warning. May be corrosive to metals. |

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Technical Guidelines

To obtain reliable and reproducible results, the operator should carefully read this entire manual and fully understand all aspects of each assay step before running the assay. The following notes should be reviewed and understood before the assay is set up.

- FOR RESEARCH USE ONLY. NOT FOR USE IN DIAGNOSTIC PROCEDURES.
- Do not use beyond the expiration date on the label.
- Do not mix or substitute reagents with those from other lots or sources.
- The Antibody-Immobilized Beads are light sensitive and must be protected from light at all times. Cover the assay plate containing beads with opaque plate lid or aluminum foil during all incubation steps.
- It is important to allow all reagents to warm to room temperature (20-25 °C) before use in the assay.
- Incomplete washing can adversely affect the assay outcome. All washing must be performed with the Wash Buffer provided.
- The standards prepared by serial dilution must be used within 1 hour of preparation. Discard any unused standards except the standard stock which may be stored at ≤ -20 °C for 1 month and at ≤ -80 °C for greater than one month.
- If samples fall outside the dynamic range of the assay, further dilute the samples with the appropriate diluent and repeat the assay.
- During the preparation of the standard curve, make certain to mix the higher concentration well before making the next dilution. Use a new tip with each dilution.
- The plate should be read immediately after the assay is finished. If, however, the plate cannot be read immediately, seal the plate, cover with aluminum foil or an opaque lid, and store the plate at 2-8 °C for up to 24 hours. Prior to reading, agitate the plate on the plate shaker at room temperature for 10 minutes. Delay in reading a plate may result in decreased sensitivity for some analytes.
- The titer plate shaker should be set at a speed to provide maximum orbital mixing without splashing of liquid outside the wells. For the recommended plate shaker, this would be a setting of 5-7 which is approximately 500-800 rpm.
- Ensure that the needle probe is clean. This may be achieved by sonication and/or alcohol flushes.
- When reading the assay on the Luminex[®] 200™ instrument, adjust probe height according to the protocols recommended by Luminex[®] to the kit solid plate or to the recommended filter plates using 3 alignment discs. When reading the assay on the MAGPIX[®] instrument, adjust probe height according to the protocols recommended by Luminex[®] to the kit solid plate or to the recommended filter plates using 2 alignment discs. When reading the assay on the FLEXMAP 3D[®] instrument, adjust probe height according to the protocols recommended by Luminex[®] to the kit solid plate using 1 alignment disc.
- For the FLEXMAP 3D[®] instrument, when using the solid plate in the kit, the final suspension should be in 150 μ L and 75 μ L should be aspirated.
- For the xMAP[®] INTELLIFLEX instrument, adjust probe height based on the type of plate you are using, place an alignment disk or an alignment sphere in the well according to the protocol recommended by Luminex[®].

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- For cell culture supernatants or tissue extraction, use the culture or extraction medium as the matrix solution in background, standard curve and control wells. If samples are diluted in assay buffer, use the assay buffer as matrix.
- For serum/plasma samples that require further dilution beyond 1:30, use the Serum Matrix provided in the kit.
- For cell/tissue homogenate, the final cell or tissue homogenate should be prepared in a buffer that has a neutral pH, contains minimal detergents or strong denaturing detergents, and has an ionic strength close to physiological concentration. Avoid debris, lipids, and cell/tissue aggregates. Centrifuge samples before use.
- Vortex all reagents well before adding to plate.

Sample Collection and Storage

Preparation of Serum Samples

- Allow the blood to clot for at least 30 minutes before centrifugation for 10 minutes at 1000 x *g*. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C.
- Avoid multiple (< 2) freeze/thaw cycles.
- When using frozen samples, it is recommended to thaw the samples completely, mix well by vortexing and centrifuge prior to use in the assay to remove particulates.
- See Preparation of Reagents Step E for detailed sample treatment information before setting up the assay.

Preparation of Plasma Samples

- Plasma collection using EDTA as an anti-coagulant is recommended. Centrifuge for 10 minutes at 1000 x *g* within 30 minutes of blood collection. Remove plasma and assay immediately or aliquot and store samples at ≤ -20 °C.
- Avoid multiple (> 2) freeze/thaw cycles.
- When using frozen samples, it is recommended to thaw the samples completely, mix well by vortexing and centrifuge prior to use in the assay to remove particulates.
- See Preparation of Reagents Step E for detailed sample treatment information before setting up the assay.

Preparation of Tissue Culture Supernatant

- Centrifuge the sample to remove debris and assay immediately or aliquot and store samples at ≤ -20 °C.
- Avoid multiple (> 2) freeze/thaw cycles.
- Tissue culture supernatant may require a dilution with an appropriate control medium prior to assay. Tissue/cell extracts should be done in neutral buffers containing reagents and conditions that do not interfere with assay performance. Excess concentrations of detergent, salt, denaturants, high or low pH, etc. will negatively affect the assay. Organic solvents should be avoided. The tissue/cell extract samples should be free of particles such as cells or tissue debris.

- See preparation of Reagents Step E for detailed sample treatment information before setting up the assay.

NOTE:

- A maximum of 25 μL per well of diluted serum or plasma can be used. Tissue culture or other media may also be used.
- All samples must be stored in polypropylene tubes. **DO NOT STORE SAMPLES IN GLASS.**
- Avoid debris, lipids and cells when using samples with gross hemolysis or lipemia.
- Care must be taken when using heparin as an anti-coagulant since an excess of heparin will provide falsely high values. Use no more than 10 IU heparin per mL of blood collected.

Preparation of Reagents for Immunoassay

Preparation of Antibody-Immobilized Beads

Sonicate the antibody-bead bottle 30 seconds and then vortex for 1 minute before use. Unused portion may be stored at 2-8 °C for up to six months.

Preparation of Quality Controls

Before use, reconstitute Quality Control 1 and Quality Control 2 with 250 μL **Assay Buffer**. Invert the vial several times to mix and vortex. Allow the vial to sit for 5-10 minutes. Transfer the reconstituted Quality Control 1 and Quality Control 2 into two polypropylene microfuge tubes. Unused portion may be stored at ≤ 20 °C for up to one month.

Preparation of Wash Buffer

Bring the 10X Wash Buffer to room temperature and mix to bring all salts into solution. Dilute 60 mL of 10X Wash Buffer with 540 mL deionized water. Store unused portion at 2-8 °C for up to one month.

Preparation of Serum Matrix

This step is required for serum or plasma samples only.

Add 1.0 mL of deionized water and 4.0 mL of Assay Buffer to the bottle containing the lyophilized Serum Matrix, mix and let it set for at least 10 min at room temperature to allow complete reconstitution. Mix well. In a separate tube, add 0.1 mL of the reconstituted Serum Matrix to 0.5 mL Assay Buffer for a final dilution of 1:30. Mix well. This is the working Serum Matrix to be used for the Standards and Controls when measuring serum/plasma samples.

Treatment of Samples

For measuring active TGF β 1, it is necessary to treat serum/plasma samples and tissue/cell culture samples prior to the assay.

Note: Tissue/cell culture samples may or may not require dilution, depending on the sample concentration of TGF β 1.

Serum/Plasma Samples

1. Centrifuge samples to remove debris and excess lipids.
2. Dilute serum/plasma samples by adding 1 part of serum/plasma to 4 parts of Sample Diluent (for example, 20 μ L serum and 80 μ L Cat. No. LTGF-SD).
3. Add 2.0 μ L of 1.0 N HCl to each 25 μ L of the 1:5 diluted samples. Make sure sample pH drops below 3.0. After mixing, acidified samples should be moderately shaken at room temperature for 15 min or incubated with no shaking for 1 hour at room temperature.
4. Acid-treated serum/plasma samples should be further diluted 1:6 using Assay Buffer (Cat. No. L-AB1) as the diluent for a final sample dilution of 1:30 immediately prior to addition to sample wells.

Tissue/Cell Culture Samples Requiring Dilution

1. Centrifuge samples to remove debris and excess lipids.
2. Dilute tissue/cell culture samples with Sample Diluent. The dilution factor should be determined by the user. For example, adding 1 part of sample to 4 parts of Sample Diluent (for example, 20 μ L sample and 80 μ L Cat. No. LTGF-SD) for 1:5 dilution.
3. Add 2.0 μ L of 1.0 N HCl to each 25 μ L of the 1:5 diluted culture medium samples. Make sure sample pH drops below 3.0. After mixing, acidified samples should be moderately shaken at room temperature for 15 min or incubated with no shaking for 1 hour at room temperature.
4. The acid-treated tissue/cell culture samples should be neutralized by 1.0 N NaOH prior to addition to sample wells. The volume of 1.0 N NaOH required is approximately similar to the volume of 1.0 N HCl used. But actual volume of 1.0 N NaOH required to achieve neutral pH may vary, depending on initial sample pH and buffering capacity of the samples.

Tissue/Cell Culture Samples Requiring NO DILUTION

1. Centrifuge samples to remove debris and excess lipids.
2. Add 2.0 μ L of 1.0 N HCl to each 25 μ L of the culture medium samples. Make sure sample pH drops below 3.0. After mixing, acidified samples should be moderately shaken at room temperature for 15 min or incubated with no shaking for 1 hour at room temperature.
3. The acid-treated tissue/cell culture samples should be neutralized by 1.0 N NaOH prior to addition to sample wells. The volume of 1.0 N NaOH required is approximately similar to the volume of 1.0 N HCl used. But actual volume of 1.0 N NaOH required to achieve neutral pH may vary, depending on initial sample pH and buffering capacity of the samples.

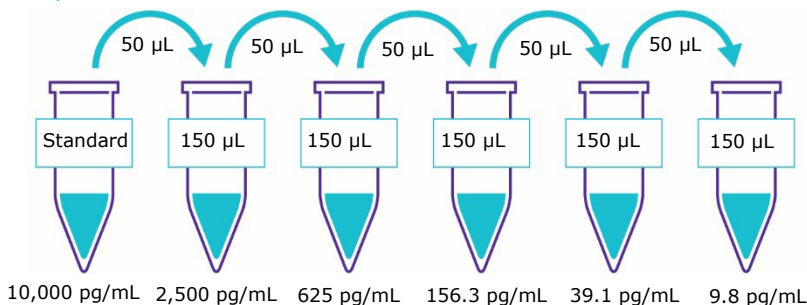
Preparation of TGFβ1 Standard

1. Prior to use, reconstitute the TGFβ1 Standard with 250 μL Assay Buffer to give a 10,000 pg/mL concentration of standard. Invert the vial several times to mix. Vortex the vial for 10 seconds. Allow the vial to sit for 5-10 minutes. Transfer the reconstituted standard to a polypropylene microfuge tube. This will be used as the 10,000 pg/mL standard; the unused portion may be stored at ≤ -20 °C for up to one month.
2. Preparation of Working Standards
Label five polypropylene microfuge tubes 2500, 625, 156.3, 39.1, and 9.8 pg/mL. Add 150 μL of the Assay Buffer to each of the five tubes. Prepare serial dilutions by adding 50 μL of the 10,000 pg/mL reconstituted standard to the 2500 pg/mL tube, mix well and transfer 50 μL of the 2500 pg/mL standard to the 625 pg/mL tube, mix well and transfer 50 μL of the 625 pg/mL standard to the 156.3 pg/mL tube, mix well and transfer 50 μL of the 156.3 pg/mL standard to the 39.1 pg/mL tube, mix well and transfer 50 μL of the 39.1 pg/mL standard to the 9.8 pg/mL tube and mix well. The 0 pg/mL standard (Background) will be the Assay Buffer.

| Standard Concentration (pg/mL) | Add Assay Buffer (μL) | Add Standard (volume) |
|--------------------------------|-----------------------|-----------------------|
| 10,000 | 250 | 0 |

| Standard Concentration (pg/mL) | Add Assay Buffer (μL) | Add Standard (volume) |
|--------------------------------|-----------------------|-----------------------|
| 2500 | 150 | 50 μL of 10,000 pg/mL |
| 625 | 150 | 50 μL of 2500 pg/mL |
| 156.3 | 150 | 50 μL of 625 pg/mL |
| 39.1 | 150 | 50 μL of 156.3 pg/mL |
| 9.8 | 150 | 50 μL of 39.1 pg/mL |

Preparation of Standards



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Immunoassay Procedure

- Prior to beginning this assay, it is imperative to read this protocol completely and to thoroughly understand the Technical Guidelines.
 - Allow all reagents to warm to room temperature (20-25 °C) before use in the assay.
 - Diagram the placement of Standards [0 (Background), 9.8, 39.1, 156.3, 625, 2500, 10,000 pg/mL], Controls 1 and 2, and Samples on Well Map Worksheet in a vertical configuration. (**Note:** Most instruments will only read the 96-well plate vertically by default.) It is recommended to run the assay in duplicate.
 - If using a filter plate, set the filter plate on a plate holder at all times during reagent dispensing and incubation steps so that the bottom of the plate does not touch any surface.
1. Add 200 μL of Assay Buffer into each well of the plate. Seal and mix on a plate shaker for 10 minutes at room temperature (20-25 °C).
 2. Decant Assay Buffer and remove the residual amount from all wells by inverting the plate and tapping it smartly onto absorbent towels several times.
 3. Add 25 μL of each Standard or Control into the appropriate wells. Assay Buffer should be used for 0 pg/mL standard (Background).
 4. Add 25 μL of Assay Buffer to the sample wells.
 5. Add 25 μL of appropriate matrix solution to the background, standards, and control wells. When assaying tissue culture samples, use identical control medium (provided by users) for Background, Standard and Quality Control wells. For serum/plasma samples, use the Serum Matrix (see Preparation of Reagents Step D.) for Background, Standard, and Quality Control wells.
 6. Add 25 μL of treated sample into the appropriate wells (see Preparation of Reagents Section Step E.).
 7. Vortex Mixing Bottle and add 25 μL of the Mixed Beads to each well. (**Note:** During addition of Beads, shake bead bottle intermittently to avoid settling.)
 8. Seal the plate with a plate sealer. Wrap the plate with foil and incubate with agitation on a plate shaker for 2 hours at room temperature (20-25 °C) or overnight (16-18 hours) at 4 °C.

Add 200 μL Assay Buffer



Shake 10 min, RT
Decant

- Add 25 μL Standard or Control to appropriate wells
- Add 25 μL Assay Buffer to background and sample wells
- Add 25 μL appropriate matrix solution to background, standards, and control wells
- Add 25 μL treated Samples to sample wells
- Add 25 μL Beads to each well



Incubate 2 hours at RT or overnight at 4 °C with shaking

9. Gently remove well contents and wash plate 2 times following instructions listed in the Plate Washing section.

10. Add 25 μL of Detection Antibodies into each well.

Note: Allow the Detection Antibodies to warm to room temperature prior to addition.

11. Seal, cover with foil and incubate with agitation on a plate shaker for 1 hour at room temperature (20-25 $^{\circ}\text{C}$). DO NOT ASPIRATE AFTER INCUBATION.

12. Add 25 μL Streptavidin-Phycoerythrin to each well containing the 25 μL of Detection Antibodies.

13. Seal, cover with foil and incubate with agitation on a plate shaker for 30 minutes at room temperature (20-25 $^{\circ}\text{C}$).

14. Gently remove well contents and wash plate 2 times following instructions listed in the Plate Washing section.

15. Add 100 μL of Sheath Fluid PLUS (or Drive Fluid PLUS if using MAGPIX[®]) to all wells. Resuspend the beads on a plate shaker for 5 minutes.

16. Run plate on Luminex[®] 200™, HTS, FLEXMAP 3D[®], MAGPIX[®] instrument with xPONENT[®] software or xMAP[®] INTELLIFLEX instrument with INTELLIFLEX software.

17. Save and analyze the Median Fluorescent Intensity (MFI) data using a 5-parameter logistic or spline curve-fitting method for calculating analyte concentrations in samples.

Note: For diluted samples, multiply the calculated concentration by the dilution factor.



Remove well contents and wash 2X with 200 μL Wash Buffer



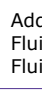
Incubate 1 hour at RT

Do Not Aspirate

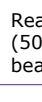


Incubate for 30 minutes at RT

Remove well contents and wash 2X with 200 μL Wash Buffer



Add 100 μL Sheath Fluid PLUS or Drive Fluid PLUS per well



Read on Luminex[®] (50 μL , 50 beads per bead set)

Plate Washing

Solid Plate

If using a solid plate, use either a handheld magnet or magnetic plate washer.

- Handheld magnet (Cat. No. 40-285)
Rest plate on magnet for 60 seconds to allow complete settling of magnetic beads. Remove well contents by gently decanting the plate in an appropriate waste receptacle and gently tapping on absorbent pads to remove residual liquid. Wash plate with 200 μ L of Wash Buffer by removing plate from magnet, adding Wash Buffer, shaking for 30 seconds, reattaching to magnet, letting beads settle for 60 seconds and removing well contents as previously described after each wash. Repeat wash steps as recommended in Assay Procedure.
- Magnetic plate washer (Cat. No. 40-094, 40-095, 40-096 and 40-097)
Please refer to specific automatic plate washer manual for appropriate equipment settings. Please note that after the final aspiration, there will be approximately 25 μ L of residual wash buffer in each well. This is expected when using the BioTek® plate washer and this volume does not need to be aspirated from the plate.

If using an automatic plate washer other than BioTek® 405 LS or 405 TS, please refer to the manufacturer's recommendations for programming instructions.

Filter Plate (Cat. No. MX-PLATE)

If using a filter plate, use a vacuum filtration manifold to remove well contents. Wash plate with 200 μ L/well of Wash Buffer, removing Wash Buffer by vacuum filtration after each wash. Repeat wash steps as recommended in the Assay Procedure.

Equipment Settings

Luminex® 200™, HTS, FLEXMAP 3D®, MAGPIX® instruments with xPONENT® software and xMAP® INTELLIFLEX instrument with INTELLIFLEX software:

These specifications are for the above listed instruments and software. Luminex® instruments with other software (for example, MasterPlex®, StarStation, LiquiChip, Bio-Plex® Manager™, LABScan™100) would need to follow instrument instructions for gate settings and additional specifications from the vendors for reading Luminex® magnetic beads.

For magnetic bead assays, each instrument must be calibrated and performance verified with the indicated calibration and verification kits.

| Instrument | Calibration Kit | Verification Kit |
|-----------------------|--|--|
| Luminex® 200™ and HTS | xPONENT® 3.1 compatible Calibration Kit (Cat. No. LX2RCAL-K25) | Performance Verification Kit (Cat. No. LX2RPVER-K25) |
| FLEXMAP 3D® | FLEXMAP 3D® Calibrator Kit (Cat. No. F3DCAL-K25) | FLEXMAP 3D® Performance Verification Kit (Cat. No. F3DPVER-K25) |
| xMAP® INTELLIFLEX | xMAP® INTELLIFLEX Calibration Kit (Cat. No. IFX-CAL-K20) | xMAP® INTELLIFLEX Performance Verification Kit (Cat. No. IFX-PVER-K20) |
| MAGPIX® | MAGPIX® Calibration Kit (Cat. No. MPXCAL-K25) | MAGPIX® Performance Verification Kit (Cat. No. MPXPVER-K25) |

NOTE: When setting up a Protocol using the xPONENT® software, you must select MagPlex as the Bead Type in the Acquisition settings.

NOTE: These assays cannot be run on any instruments using Luminex® IS 2.3 or Luminex® 1.7 software.

The Luminex® probe height must be adjusted to the plate provided in the kit. Please use Cat. No. MAG-PLATE, if additional plates are required for this purpose.

| | |
|---------------|---------------------|
| Events | 50, per bead |
| Sample Size | 50 µL |
| Gate Settings | 8,000 to 15,000 |
| Reporter Gain | Default (low PMT) |
| Time Out | 60 seconds |
| Bead Set | TGFβ1 18 |

Quality Controls

The ranges for each analyte in Quality Control 1 and 2 are provided on the card insert or can be located at our website SigmaAldrich.com using the catalogue number as the keyword.

Assay Characteristics

Cross-Reactivity

There was no or negligible cross-reactivity between the antibodies for an analyte and any of the other analytes in this panel.

- In addition to human TGFβ1, the assay can detect TGFβ1 in serum of the following: horse, rabbit, mouse, rat, mini pig, canine, hamster, guinea pig, rhesus monkey and cynomolgous monkey. However, the exact amount of cross reactivity has not been determined.

Assay Sensitivities (minimum detectable concentrations, pg/mL)

Minimum Detectable Concentration (MinDC) is calculated using MILLIPLEX® Analyst 5.1. It measures the true limits of detection for an assay by mathematically determining what the empirical MinDC would be if an infinite number of standard concentrations were run for the assay under the same conditions.

| Analyte | Overnight Protocol (n=5 Assays) | | 2 Hour Protocol (n = 2 Assays) | |
|---------|------------------------------------|----------------------|-----------------------------------|----------------------|
| | MinDC (pg/mL) | MinDC+2SD (pg/mL) | MinDC (pg/mL) | MinDC+2SD (pg/mL) |
| TGFβ1 | 6.0 | 11.4 | 9.3 | 11.9 |

Precision

Intra-assay precision is generated from the mean of the %CV's from **8** reportable results across two different concentrations of analytes in a single assay. Inter-assay precision is generated from the mean of the %CV's across two different concentrations of analytes across 5 different assays.

| Analyte | Overnight Protocol | |
|---------|--------------------|-----------------|
| | Intra-assay %CV | Inter-assay %CV |
| TGFβ1 | 7.2 | 4.6 |

Accuracy

Spike Recovery: The data represent mean percent recovery of spiked standards ranging from low, medium, and high concentration in serum matrices (n=5).

| Analyte | Overnight Protocol |
|---------|----------------------------|
| | % Recovery in Serum Matrix |
| TGFβ1 | 99.5 |

Troubleshooting

| Problem | Probable Cause | Solution |
|-------------------------|---|---|
| Insufficient bead count | Plate washer aspirate height set too low | Adjust aspiration height according to manufacturers' instructions. |
| | Bead mix prepared inappropriately | Sonicate bead vials and vortex just prior to adding to bead mix bottle according to protocol. Agitate bead mix intermittently in reservoir while pipetting this into the plate. |
| | Samples cause interference due to particulate matter or viscosity | See above. Also sample probe may need to be cleaned with alcohol flushes, back flushes and washes; or if needed, probe should be removed and sonicated. |
| | Probe height not adjusted correctly | When reading the assay on the Luminex® 200™ instrument, adjust probe height to the kit solid plate or to the recommended filter plates using 3 alignment discs. When reading the assay on the MAGPIX® instrument, adjust probe height to the kit solid plate or to the recommended filter plates using 2 alignment discs. When reading the assay on the FLEXMAP 3D® instrument, adjust probe height to the kit solid plate using 1 alignment disc. For the FLEXMAP 3D® instrument, when using the solid plate in the kit, the final suspension should be in 150 µL and 75 µL should be aspirated. When reading the assay on the xMAP® INTELLIFLEX instrument, adjust probe height based on the type of plate you are using, place an alignment disk or an alignment sphere in the well according to the protocol recommended by Luminex®. |
| Background is too high | Background wells were contaminated | Avoid cross-well contamination by using sealer appropriately and pipetting with multichannel pipettes without touching reagent in plate. |
| | Matrix used has endogenous analyte or interference | Check matrix ingredients for cross-reacting components (for example, interleukin modified tissue culture medium). |
| | Insufficient washes | Increase number of washes. |

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| Problem | Probable Cause | Solution |
|---|---|---|
| Beads not in region or gate | Luminex® instrument not calibrated correctly or recently | Calibrate Luminex® instrument based on manufacturer's instructions, at least once a week or if temperature has changed by > 3 °C. |
| | Gate settings not adjusted correctly | Some Luminex® instruments (for example, Bio-Plex®) require different gate settings than those described in the kit protocol. Use instrument default settings. |
| | Wrong bead regions in protocol template | Check kit protocol for correct bead regions or analyte selection. |
| | Incorrect sample type used | Samples containing organic solvents or if highly viscous should be diluted or dialyzed as required. |
| | Instrument not washed or primed | Prime the Luminex® instrument 4 times to rid it of air bubbles, wash 4 times with sheath fluid or water if there is any remnant alcohol or sanitizing liquid. |
| | Beads were exposed to light | Keep plate and bead mix covered with dark lid or aluminum foil during all incubation steps. |
| Signal for whole plate is same as background | Incorrect or no Detection Antibody was added | Add appropriate Detection Antibody and continue. |
| | Streptavidin-Phycoerythrin was not added | Add Streptavidin-Phycoerythrin according to protocol. If Detection Antibody has already been removed, sensitivity may be low. |
| Low signal for standard curve | Detection Antibody may have been removed prior to adding Streptavidin-Phycoerythrin | May need to repeat assay if desired sensitivity not achieved. |
| | Incubations done at inappropriate temperatures, timings or agitation | Assay conditions need to be checked. |
| Signals too high, standard curves are saturated | Calibration target value set too high | With some Luminex® instruments (for example, Bio-Plex®) default target setting for RP1 calibrator is set at high PMT. Use low target value for calibration and reanalyze plate. |
| | Plate incubation was too long with standard curve and samples | Use shorter incubation time. |

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| Problem | Probable Cause | Solution |
|--|---|---|
| Sample readings are out of range | Samples contain no or below detectable levels of analyte | If below detectable levels, it may be possible to use higher sample volume. Check with technical support for appropriate protocol modifications. |
| | Samples contain analyte concentrations higher than highest standard point | Samples may require dilution and reanalysis for just that particular analyte. |
| | Standard curve was saturated at higher end of curve | See above. |
| High variation in samples and/or standards | Multichannel pipette may not be calibrated | Calibrate pipettes. |
| | Plate washing was not uniform | Confirm all reagents are removed completely in all wash steps. |
| | Samples may have high particulate matter or other interfering substances | See above. |
| | Plate agitation was insufficient | Plate should be agitated during all incubation steps using an orbital plate shaker at a speed where beads are in constant motion without causing splashing. |
| | Cross-well contamination | Check when reusing plate sealer that no reagent has touched sealer. Care should be taken when using same pipette tips that are used for reagent additions and that pipette tip does not touch reagent in plate. |

FOR FILTER PLATES ONLY

| Problem | Probable Cause | Solution |
|------------------------------|---|--|
| Filter plate will not vacuum | Vacuum pressure is insufficient | Increase vacuum pressure such that 0.2 mL buffer can be suctioned in 3-5 seconds. |
| | Samples have insoluble particles | Centrifuge samples just prior to assay set-up and use supernatant. |
| | High lipid concentration | After centrifugation, remove lipid layer and use supernatant. |
| Plate leaked | Vacuum pressure too high | Adjust vacuum pressure such that 0.2 mL buffer can be suctioned in 3-5 seconds. May need to transfer contents to a new (blocked) plate and continue. |
| | Plate set directly on table or absorbent towels during incubations or reagent additions | Set plate on plate holder or raised edge so bottom of filter is not touching any surface. |
| | Insufficient blotting of filter plate bottom causing wicking | Blot the bottom of the filter plate well with absorbent towels after each wash step. |
| | Pipette touching plate filter during additions | Pipette to the side of plate. |
| | Probe height not adjusted correctly | Adjust probe to 3 alignment discs in well H6. |
| | Sample too viscous | May need to dilute sample. |

Product Ordering

| Replacement Reagents | Cat. No. |
|--|------------------|
| TGF β 1 Standard | LTGF-8064 |
| TGF β 1 Quality Controls | LTGF-6064 |
| TGF β 1 Detection Antibodies | LTGF-1064 |
| Sample Diluent | LTGF-SD |
| Streptavidin-Phycoerythrin | L-SAPE9 |
| Serum Matrix | LMC-SD |
| 1.0 N HCl | L-HCL |
| 1.0 N NaOH | L-NAOH |
| Assay Buffer | L-AB1 |
| Wash Buffer | L-WB |
| Set of two 96-Well Filter Plates with Sealers | MAG-PLATE |
| TGF β 1 Single Plex Magnetic Bead Kit – BULK PACKAGING | TGFBMAG-64K-01BK |

Antibody-Immobilized Magnetic Beads

| Analyte | Bead No. | Cat. No. |
|----------------|-----------------|-----------------|
| TGF β 1 | 18 | TGFB1-MAG |

Well Map

| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|--|----------------------------|-----------------|---|---|---|---|---|---|----|----|----|
| A | 0 pg/mL Standard (Background) | 625 pg/mL Standard | QC 2 Control | | | | | | | | | |
| B | 0 pg/mL Standard (Background) | 625 pg/mL Standard | QC 2 Control | | | | | | | | | |
| C | 9.8 pg/mL Standard | 2500 pg/mL Standard | Sample 1 | | | | | | | | | |
| D | 9.8 pg/mL Standard | 2500 pg/mL Standard | Sample 1 | | | | | | | | | |
| E | 39.1 pg/mL Standard | 10000 pg/mL Standard | Sample 2 | | | | | | | | | |
| F | 39.1 pg/mL Standard | 10000 pg/mL Standard | Sample 2 | | | | | | | | | |
| G | 156.3 pg/mL Standard | QC 1 Control | Etc. | | | | | | | | | |
| H | 156.3 pg/mL Standard | QC 1 Control | | | | | | | | | | |

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